

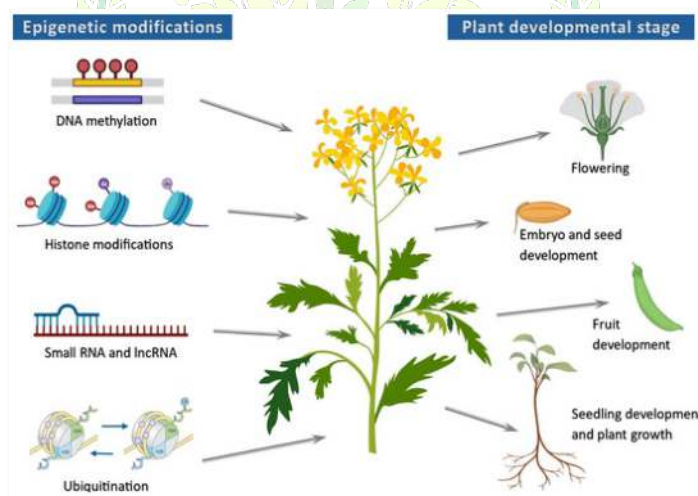
# Epigenetic Modulation of Plant Disease Resistance

ARTICLE ID: 0316

Poulami Basak<sup>1\*</sup>, Jenia Roy<sup>2</sup>, Dipsikha Mondal<sup>3</sup><sup>1</sup>Division of Plant Pathology, ICAR–Indian Agricultural Research Institute (IARI), New Delhi 110012, India<sup>2</sup>Division of Genetics, ICAR–Indian Agricultural Research Institute (IARI), New Delhi 110012, India<sup>3</sup>Division of Agricultural Chemicals, ICAR–Indian Agricultural Research Institute (IARI), New Delhi 110012, India

Plants, being sessile organisms, are associated with disease resistance can be effectively continuously exposed to a wide range of exploited in crop-breeding programmes for the biotic and abiotic stresses. To successfully development of improved and resilient varieties.

Plants defend themselves against pathogens through sophisticated defence mechanisms that have evolved over time. Pattern-triggered immunity (PTI) is activated when pattern recognition receptors (PRRs) recognize pathogen-associated molecular patterns



repel invading pathogens, plants have evolved an intricate and highly regulated immune system that enables rapid and need-based activation of defence genes. Epigenetics refers to heritable changes in gene function that occur without alterations in the underlying DNA sequence and can be transmitted mitotically and/or meiotically. Epigenetic regulation—mediated through DNA methylation, histone modifications, and small RNAs—plays a critical role in controlling gene expression during stress responses. With advances in molecular tools, epigenome editing has recently emerged as a promising strategy for enhancing resistance against various plant stresses. Stable and heritable epigenetic regulatory mechanisms

(PAMPs) or damage-associated molecular patterns (DAMPs). In contrast, effector-triggered immunity (ETI) is initiated when plant resistance (R) proteins detect pathogen-secreted effector molecules (Jones et al., 2016). Both PTI and ETI involve extensive transcriptional reprogramming of defence-related genes; however, they differ in the magnitude and duration of downstream responses.

Many defence-associated genes are located in heterochromatic regions and are transcriptionally inaccessible under normal conditions. Activation of these genes requires coordinated action of epigenetic regulators that modulate DNA methylation patterns, chromatin structure, and histone modifications (Qi et al., 2023).

### **Types of Epigenetic Mechanisms**

Epigenetic mechanisms can be broadly classified into three major categories: DNA methylation/demethylation, histone modifications, and non-coding RNAs.

#### **DNA Methylation**

DNA methylation involves the addition of a methyl group to the 5-position of cytosine residues, catalysed by DNA methyltransferases (MTases). It is generally associated with transcriptional repression, particularly when present in promoter regions, and results in heritable methylation patterns transmitted during mitosis and meiosis. DNA methylation is regulated through both methylation (maintenance and de novo) and demethylation (active and passive) processes. Maintenance methylation preserves existing methylation patterns during DNA replication, whereas de novo methylation establishes new methylation marks on previously unmethylated regions, primarily via the RNA-directed DNA methylation (RdDM) pathway. In plants, DNA methylation is controlled by three major families of methyltransferases: maintenance methyltransferases (METs), chromomethylases (CMTs), and de novo DNA methyltransferases (DRMs).

#### **Histone Modifications**

Histones are basic proteins rich in lysine and arginine residues, which impart a positive charge and facilitate interaction with negatively charged DNA. The N-terminal tails of histones are subject to diverse post-translational modifications, including methylation, acetylation, ubiquitination, and phosphorylation. Histone methylation is among the most extensively studied modifications and can either activate or repress gene expression depending on the specific residue modified; for example, H3K4 methylation is associated with transcriptional activation, whereas H3K27 methylation is linked to gene repression. Histone acetylation neutralizes the positive charge on lysine residues, reducing histone–DNA affinity and thereby enhancing accessibility of transcriptional machinery such as transcription factors and RNA polymerase II.

#### **Non-coding RNAs**

Non-coding RNAs are RNA molecules that do not encode proteins but play crucial roles in gene regulation and genome stability. Major classes include microRNAs (miRNAs) and small interfering RNAs (siRNAs), which mediate RNA interference pathways by promoting mRNA degradation or inhibiting translation, thereby regulating gene expression during plant–pathogen interactions.

#### **Applications in Plant Disease Resistance**

Both naturally occurring epigenetic variation and artificially induced epigenetic modifications can significantly influence plant responses to pathogens. These mechanisms offer considerable potential for epigenetic breeding approaches aimed at improving

disease resistance. Selected examples of epigenetic regulators implicated in plant disease resistance are summarized in Table 1.

**Table 1. Reported use of epigenetic regulators in plant disease resistance**

| Gene name                        | Gene function   | Role in host–pathogen interaction   | Reference                   |
|----------------------------------|---|---|-----------------------------|
| <b>AeDRM2</b>                    | De novo DNA methyltransferase                             | Knockdown enhanced resistance of <i>Aegilops tauschii</i> to <i>Blumeria graminis</i> f. sp. <i>tritici</i> | Geng et al., 2019           |
| <b>Xa21G</b>                     | Resistance to <i>Xanthomonas oryzae</i> pv. <i>oryzae</i> | Demethylation using 5-azadeoxycytidine improved disease resistance  | Akimoto et al., 2007        |
| <b>IR-specific siRNA (24 nt)</b> | Transcriptional gene silencing                            | Increased methylation of MYMIV intergenic region conferred resistance in soybean                            | Yadav & Chattopadhyay, 2011 |
| <b>MeSWEET10a</b>                | Susceptibility gene in cassava                            | Targeted methylation of EBE site conferred resistance to bacterial blight                                   | Veley et al., 2023          |
| <b>SIPR1</b>                     | Defence-related gene                                      | H3K4me3 enrichment enhanced resistance to <i>Clavibacter michiganensis</i> ssp. <i>michiganensis</i>        | García-Murillo et al., 2023 |

## Conclusion

Epigenetic modification represents a promising alternative strategy for developing disease-resistant crop varieties through epi-breeding. Epigenomic variations associated with disease resistance traits can serve as molecular epigenetic markers to support selection and evaluation in breeding programmes. Furthermore, epigenetic modelling may help predict the effects of epigenomic variation on disease resistance and guide targeted epigenome engineering. Future research should focus on elucidating the precise regulatory mechanisms by which epigenetic factors are recruited to defence genes during plant immune responses.

## References

1. Akimoto, K., Katakami, H., Kim, H., Ogawa, E., Sano, C. M., Wada, Y., & Sano, H. (2007). Epigenetic inheritance in rice plants. *Annals of Botany*, **100**(2), 205–217.
2. García-Murillo, L., Valencia-Lozano, E., Priego-Ranero, N. A., Cabrera-Ponce, J. L., Duarte-Aké, F. P., Vizuet-de-Rueda, J. C., Rivera-Toro, D. M., Herrera-Ubaldo, H., de Folter, S., & Alvarez-Venegas, R.

- (2023). CRISPRa-mediated transcriptional activation of the *SlPR-1* gene in edited tomato plants. *Plant Science*, 111617.
3. Geng, S., Kong, X., Song, G., Jia, M., Guan, J., Wang, F., Qin, Z., Wu, L., Lan, X., Li, A., & Mao, L. (2019). DNA methylation dynamics during the interaction of *Aegilops tauschii* with *Blumeria graminis* f. sp. *tritici*. *New Phytologist*, **221**(2), 1023–1035.
  4. Jones, J. D. G., Vance, R. E., & Dangl, J. L. (2016). Intracellular innate immune surveillance devices in plants and animals. *Science*, **354**, 6316.
  5. Qi, Q., Hu, B., Jiang, W., Wang, Y., Yan, J., Ma, F., Guan, Q., & Xu, J. (2023). Advances in plant epigenome editing research and its application in plants. *International Journal of Molecular Sciences*, **24**(4), 3442.
  6. Veley, K. M., Elliott, K., Jensen, G., Zhong, Z., Feng, S., Yoder, M., Gilbert, K. B., Berry, J. C., Lin, Z. D., Ghoshal, B., Gallego-Bartolomé, J., Norton, J., Motomura-Wages, S., Carrington, J. C., Jacobsen, S. E., & Bart, R. (2023). Improving cassava bacterial blight resistance by editing the epigenome. *Nature Communications*, **14**, 1–12.
  7. Yadav, R. K., & Chattopadhyay, D. (2011). Enhanced viral intergenic region-specific siRNA accumulation and DNA methylation correlate with resistance against a geminivirus. *Molecular Plant–Microbe Interactions*, **24**(10), 1189–1197.

